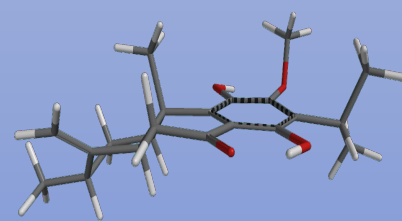


# Binding of Telomeric DNA G-Quadruplexes by Abietane Diterpene Natural Products

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## Introduction

G-quadruplexes are non-canonical higher order DNA structures formed from guanine rich sequences, made up of stacked G-tetrads stabilized by Hoogsteen base pairing and  $K^+$  ions. G-quadruplexes are overrepresented in the promoter regions of oncogenes and the 5'UTR of mRNA<sup>1</sup>. As a result, G-Quadruplexes have been implicated as targets for possible anti-cancer therapeutic agents to treat previously “undruggable” targets like the c-myc and ras oncogenes<sup>1</sup>.

The human telomeric repeat,  $[5'G_3(T_2AG_3)_3]$ , is a repeating, single stranded DNA sequence that can form G-quadruplexes. Telomerase, an enzyme expressed in ~90% of all cancers, is responsible for extending telomeric repeats, making cancer cells immortal. It has been shown that stabilization of telomeric G-quadruplexes can inhibit telomerase activity and therefore block the survival of cancer cells<sup>2</sup>.

The compounds used in the study are a group of abietane diterpene natural products from *Hyptis verticillata*, a plant native to the Caribbean and central America that has been used traditionally as an ethnomedicine that has been shown to have a range of therapeutic effects. Some of these effects include anti-microbial, anti-inflammatory, and even anti-cancer activities<sup>3</sup>. Using several biophysical techniques, we have investigated the binding characteristics of these compounds to G-quadruplex DNA as a possible rationale for their observed anti-cancer therapeutic effects.

## Experimental

### Sample Preparation

DNA G-Quadruplexes were formed in solution using a 10 mM phosphate buffer, pH 7, with 100mM  $K^+$  added. Characteristic CD spectra confirmed the presence of quadruplexes in solution.

### Circular dichroism

CD measurements were performed using a Jasco J-815 CD Spectrophotometer. For each run, the abietane solution was added to a solution of G-quadruplex DNA and the spectra was measured from 200-600nm.

### Thermal Denaturation

Tm measurements were performed using a CARY-5000 UV-Vis-NIR Spectrophotometer. Absorption spectra of G-quadruplex-abietane complex were recorded at 295 nm as the temperature was increased from 25.00°C to 95.00°C.

### Thioflavin-T Fluorescence Displacement

Fluorescent studies were performed using a PTI Quantamaster™ 40 UV-Vis Spectrofluorometer from 400-600nm. A Fluorogenic dye that binds specifically to DNA G-quadruplexes, Thioflavin-T (ThT), was used as a fluorescent indicator for the quadruplexes. A solution of ThT bound G-quadruplexes was prepared and the abietane solution was added. The solution was excited at 425 nm and fluorescence intensity was measured.

## Thioflavin Displacement

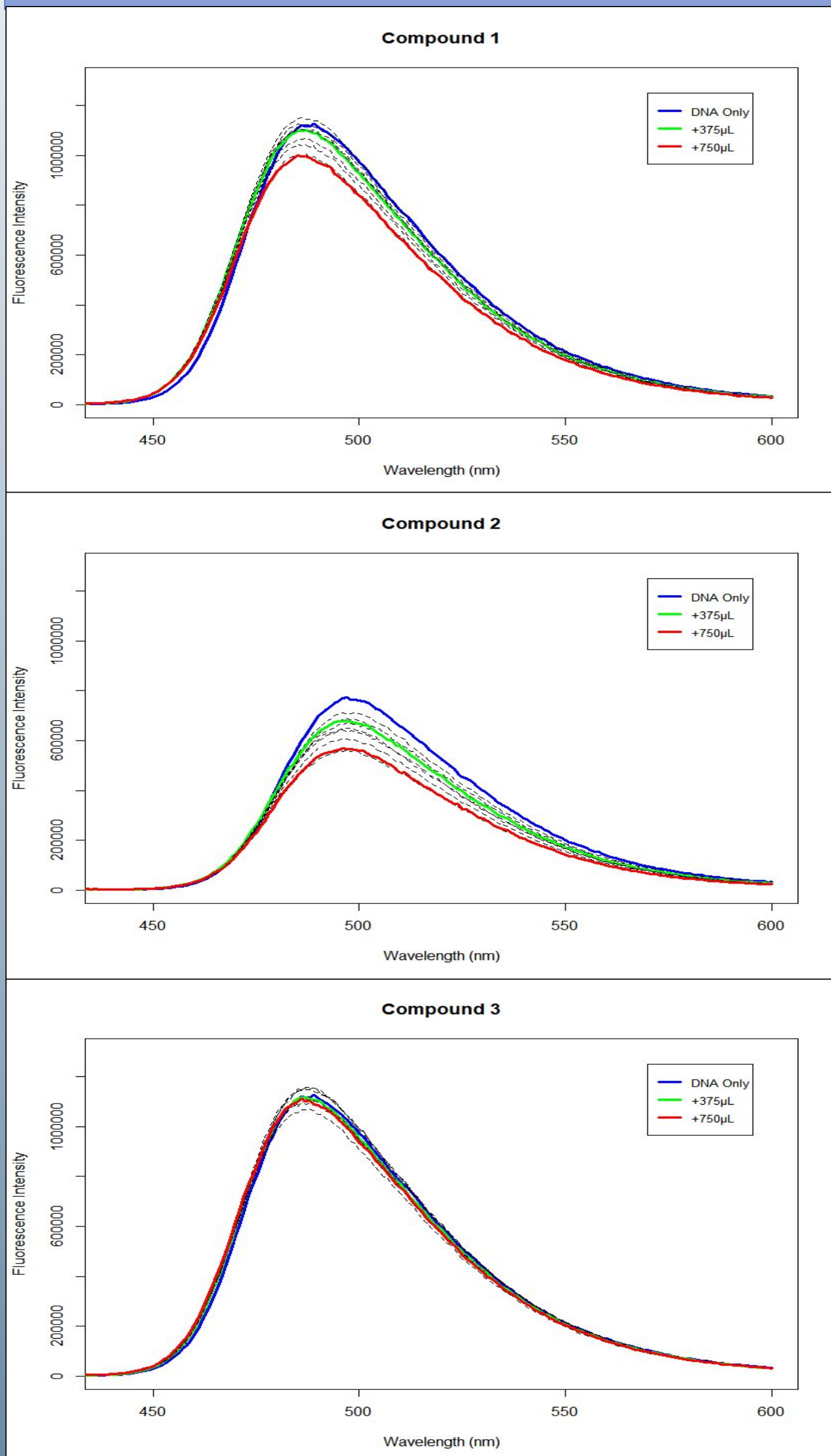


Figure 4. Thioflavin-T displacement graphs of compounds 1, 2 and 3. As the abietanes were bound, the fluorescence intensity changed base on whether or no the abietanes were able to displace Thioflavin-T from its binding site on the quadruplexes.

## Results

## Circular Dichroism

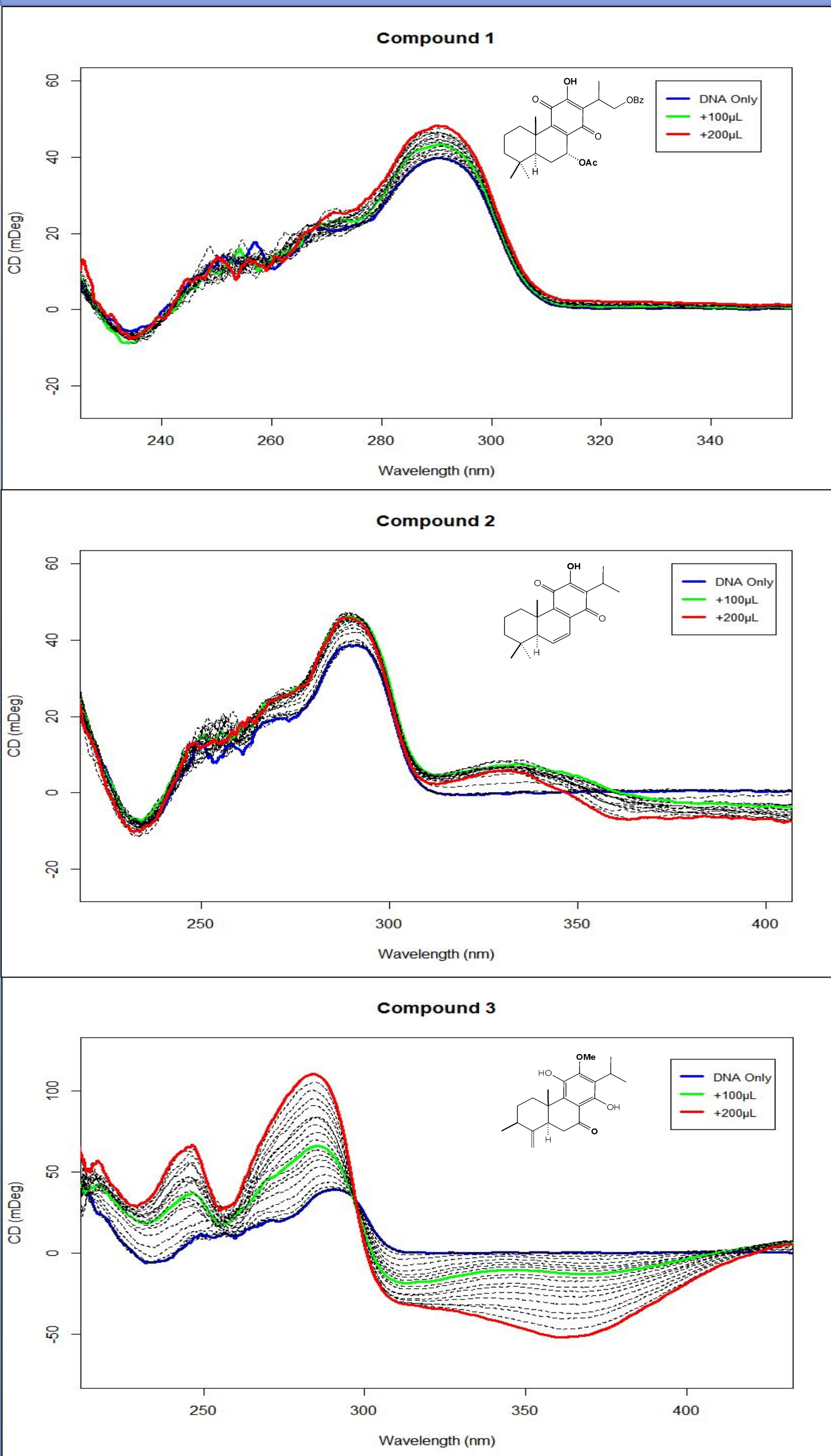


Figure 5. CD Spectra of three abietanes (compound 1, compound 2, compound 3) after being titrated into G-quadruplex solution. Structures of compounds 2 and 3 are shown.

## Thermal Denaturation

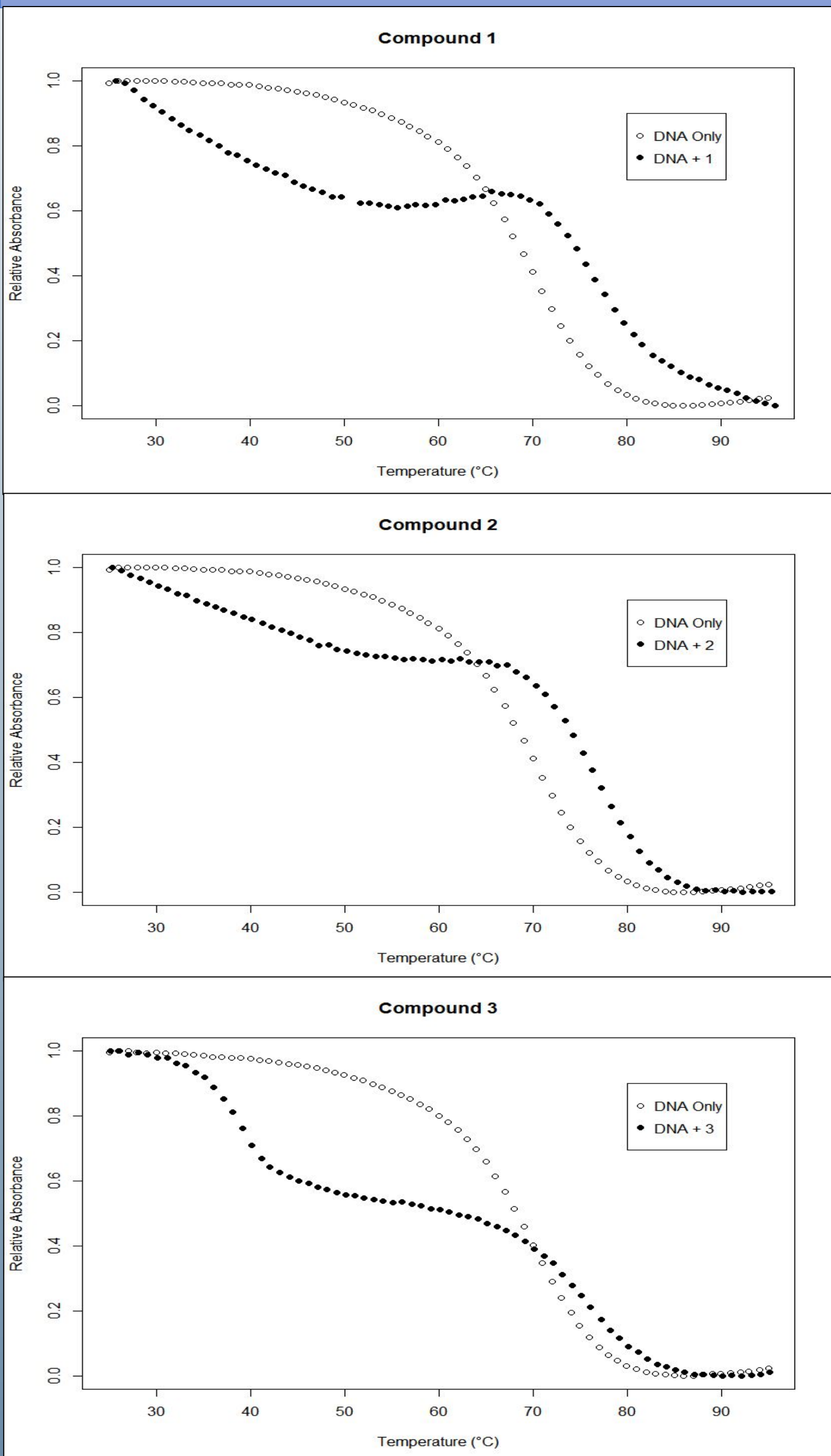


Figure 6. Thermal Denaturation plots of compounds 1, 2, and 3. Each compound was run alongside a DNA only control.

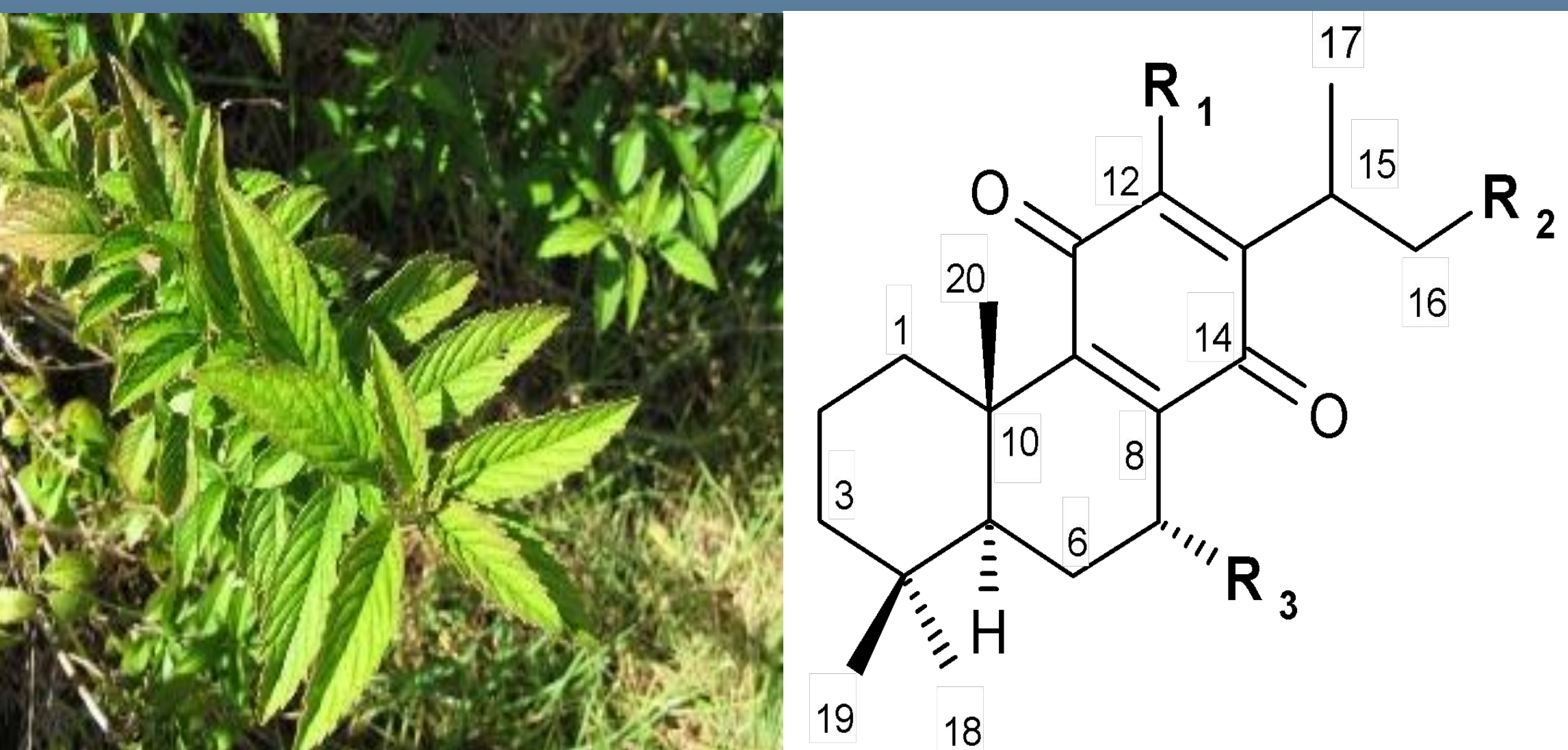


Figure 1. Left: A picture of *Hyptis verticillata*, the plant from which these compounds are derived<sup>5</sup>. Right: A generalized structure of the abietane diterpenes used.

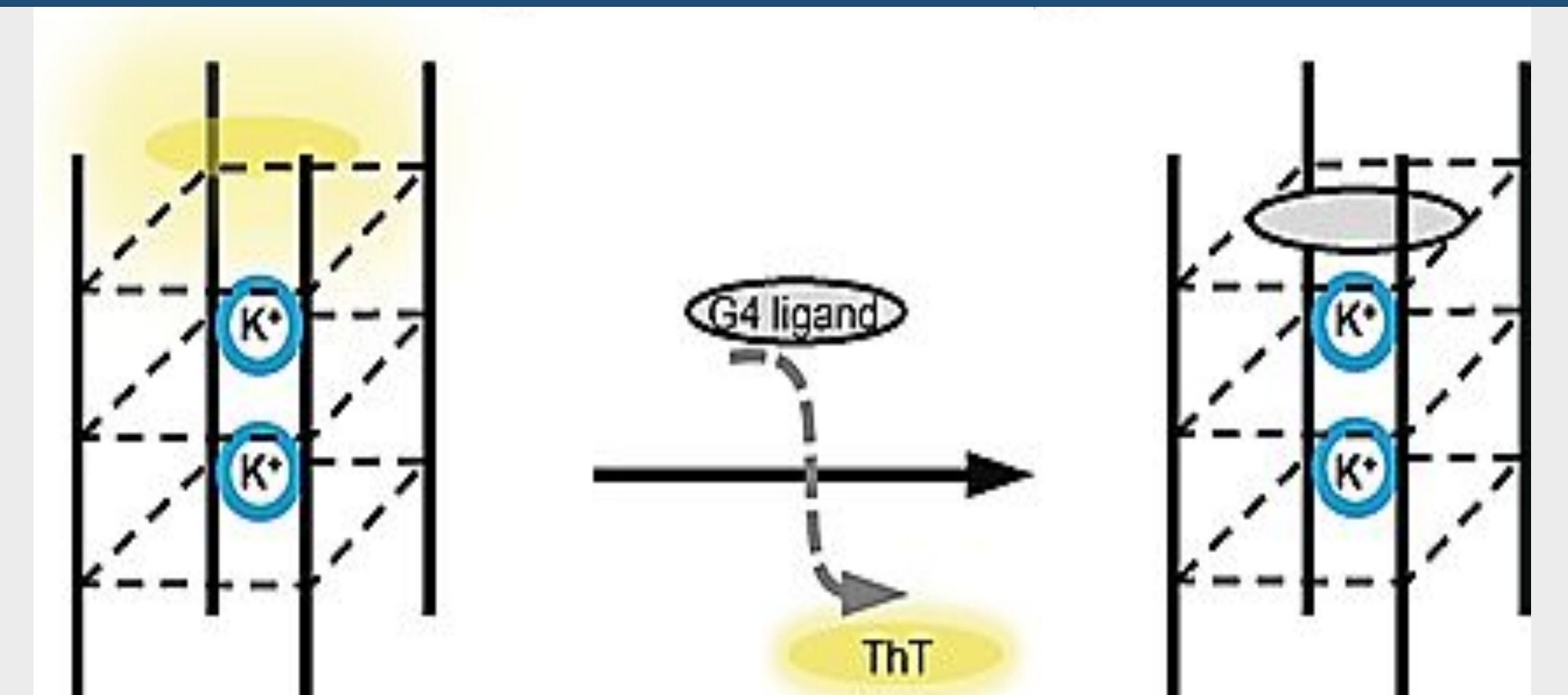


Figure 2. A simple diagram demonstrating the mechanism of the Thioflavin-T displacement assay, Adapted from Jamroskovic et al<sup>7</sup>.

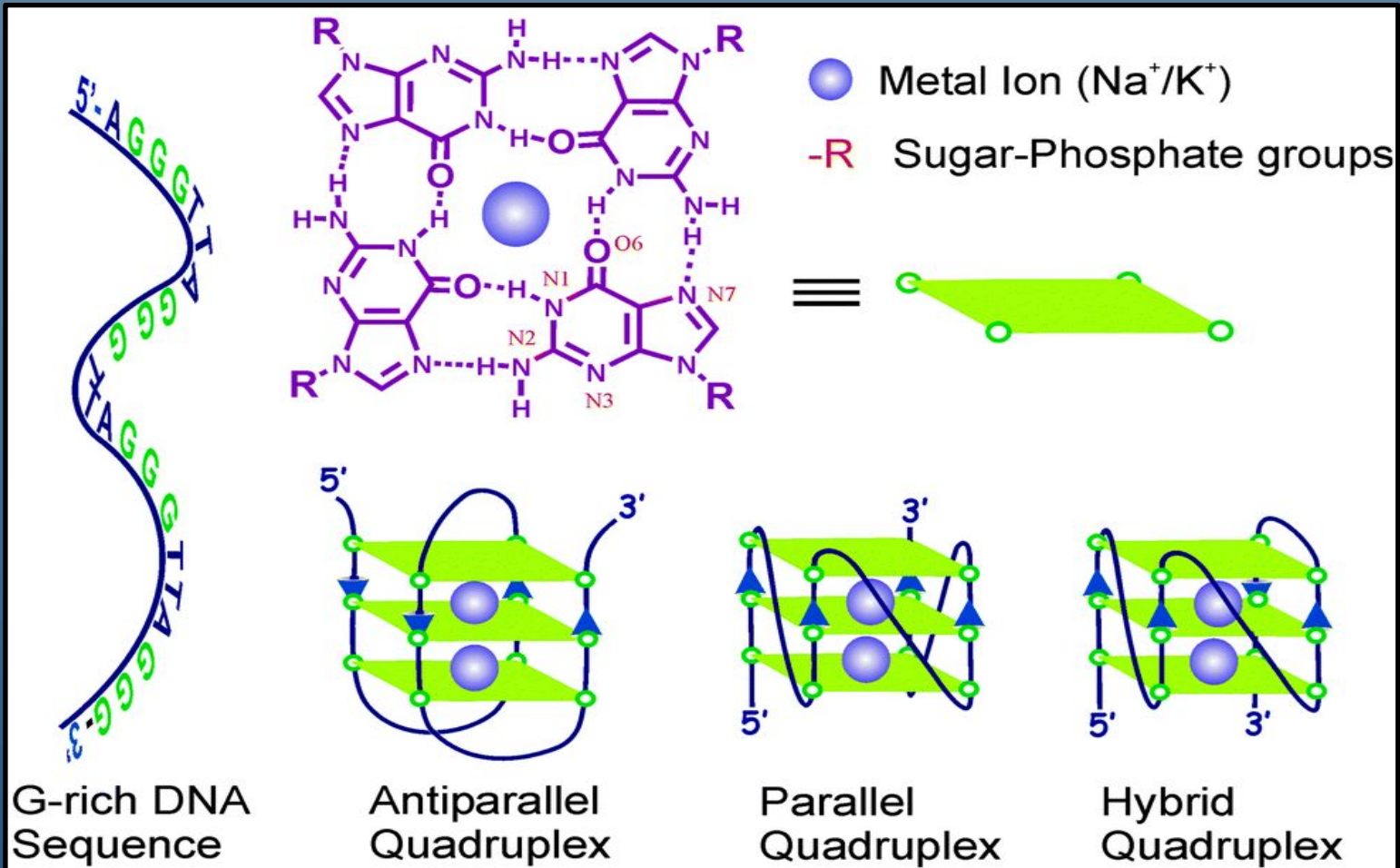


Figure 3. A simplified diagram of DNA G-quadruplexes formed from human telomeric DNA<sup>6</sup>.

Table 1.  $\Delta T_m$  values of each compound determined from the thermal denaturation studies. Apparent Stern-Volmer constants were determined for each compound via the Thioflavin-T displacement studies.

Compound	$\Delta T_m$ (°C)	Stern-Volmer Constant ( $mM^{-1}$ )
1	5.7	0.0075
2	5.4	0.0169
3	-17.86	-
4	8.1	0.0151
5	8.4	0.0118
6	8.46	0.0136
7	-	-
8	2.2	-

## Conclusions

- Displacement studies showed that 5 of the 8 abietane diterpenes interacted with the DNA-ThT complex, quenching the fluorescence.
- CD Spectra indicated that of the 8 compounds tested, compounds 1, 2, and 3 showed the most significant interactions with the telomeric G-quadruplexes.
- The unique spectra of compound 3 indicates a shift from antiparallel quadruplexes to a different form of quadruplex, although it is unclear what that form may be. The positive peaks at 295nm and negative peaks at 240nm shown by the spectra for compounds 1 and 2 indicate an increase in stabilization of the antiparallel form of G-quadruplex<sup>8</sup>.
- Thermal denaturation studies showed that the binding of all the compounds resulted in an increase in thermal stability ( $T_m$ ) at higher temperatures, after an initial structural transition. An early transition/species was most evident for 3.
- The spectra we have recorded for all compounds, specifically compound 3, is very interesting and warrants further investigation.

## References

- Balasubramanian et al. Targeting G-quadruplexes in gene promoters: a novel anticancer strategy? Nat Rev Drug Discov. 2011; 10:261-275
- Sun et al. Inhibition of human telomerase by a G-quadruplex-interactive compound. J. Med. Chem. 1997; 40:2113-2116
- Picking et al. *Hyptis verticillata* Jacq: A review of its traditional uses, phytochemistry, pharmacology and toxicology. Journal of Ethnopharmacology. 2013; 147:16-41
- Mohanty et al. Thioflavin T as an Efficient Inducer and Selective Fluorescent Sensor for the Human Telomeric G-Quadruplex DNA. Journal of the American Chemical Society. 2013; 135:367-376
- <https://regionalconservation.org/ircs/database/plants/PlantPage.asp?TXCODE=Hyptvert>
- Bhasikuttan, Mohanty. Targeting G-quadruplex Structures with extrinsic fluorogenic dyes: promising fluorescence sensors. Chem. Commun. 2015; 51:7581-7597
- Jamroskovic et al. Identification of Compounds that Selectively Stabilize Specific G-Quadruplex Structures by using a Thioflavin T-Displacement Assay as a tool. Chemistry – A European Journal. 2016; 52:18932-18943
- Dai et al. Polymorphism of human telomeric quadruplex structures. Biochimie. 2008;90(8):1172–1183

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